

**Noda Institute for Scientific Research GRANT**  
**2014 Research Grant**  
**Outline of Research Result**

# Ergothioneine production by methylotrophic bacteria

Akio TANI

*Institute of Plant Science and Resources, Okayama University*

## Research aims

*Methylobacterium* species are methylotrophic bacteria belonging to Alphaproteobacteria, and have been studied in detail as a model for bacterial methylotrophy. Recent research studies have revealed that plants emit huge amounts of methanol and that these methylotrophic bacteria are one of the major bacterial species inhabiting the plant surface.<sup>1)</sup> Furthermore, inoculation of the methylotrophic bacteria may promote plant growth.<sup>2)</sup> During our research on the mechanism of plant growth promotion, we found through metabolome analysis that one of the species accumulated ergothioneine (EGT; Fig. 1). In this study, we aimed to investigate the physiological role of EGT accumulation in the species and the feasibility of fermentative production of EGT.

## Methods

*Methylobacterium aquaticum* strain 22A was used as a model in this study.<sup>3)</sup> The strain was isolated from the bryophyte *Racomitrium japonicum*, the complete genome of which has been sequenced.<sup>4)</sup> A plasmid, pK18mobSacB, was used for generating a gene deletion mutant of this strain. HPLC was used for EGT quantification. Amino acids were analyzed with an amino acid analyzer.

## Results

In the metabolome analysis of strain 22A cells grown on methanol, EGT was found as a most prominent peak by capillary electrophoresis-mass spectrometry. A quantification method based on HPLC analysis was established. Methanol was found to be the best substrate for EGT production among other carbon sources, such as ethanol, succinate, and glucose. EGT was the most abundant amino acid

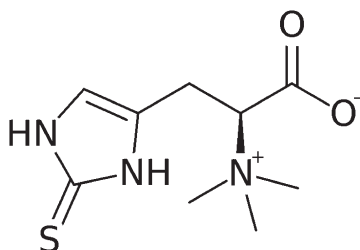


Fig. 1. Ergothioneine

accumulated in the cell compared with other amino acids (Fig. 2). We also found that most *Methylobacterium* species could synthesize EGT, with strain 22A being one of the best strains among them. The cultivation condition was optimized by changing the concentration of the nitrogen sources, iron, lanthanoid, and amino acids. As a result, the productivity reached up to 1 mg/100 mL culture medium (1.2 mg/g wet weight cells, 6.3 mg/g dry weight cells) using 2% methanol as a carbon source in one month of cultivation. The productivity overwhelmed that of *Pleurotus ostreatus* (oyster mushroom).<sup>5)</sup>

EGT biosynthesis genes were first identified as a gene cluster (*egtABCDE*) in *Mycobacterium* species.<sup>6)</sup> We found the homologous genes to be scattered among the genome of strain 22A, except for *egtB* and *egtD* that were clustered. A deletion mutant of *egtBD* was generated, which completely abolished the productivity of EGT. The mutant ( $\Delta egtBD$ ) showed more severe susceptibility to heat shock and UV irradiation and higher resistance to hydrogen peroxide than the wild type. We also found that EGT could be degraded by UV irradiation, and the intracellular EGT content decreased upon UV irradiation of the mutant cells. In addition, the mutant was more susceptible to sunlight irradiation. However, the mutant could grow on methanol, suggesting that EGT is not involved in methylotrophy.

## Conclusion

Taken together, we showed that it is possible to produce EGT from *Methylobacterium* species utilizing methanol as a carbon source. High-cell density cultivation using the

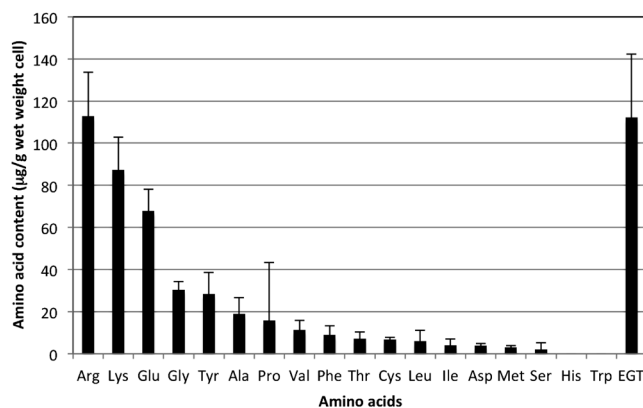


Fig. 2. Intracellular amino acids in strain 22A.

genus is possible, with a yield of 100 g dry weight cells/liter.<sup>7)</sup> We are still optimizing the production by examining the high-cell density cultivation, screening microorganisms with higher productivity, optimizing the cultivation conditions, and promoting the gene expression. In this study, we found that EGT accumulates in high amounts when the strain is grown on methanol, but EGT is not involved in methylotrophy. Since the mutant showed higher susceptibility to sunlight, EGT would appear to be an important amino acid for the genus to inhabit the plant surface.

## References

- 1) Vorholt, J. A. (2012) Microbial life in the phyllosphere. *Nat. Rev. Microbiol.* **10**: 828–840.
- 2) Abanda-Nkpwatt, D., Müsch, M., Tschiersch, J., Boettner, M., and Schwab, W. (2006) Molecular interaction between *Methylobacterium extorquens* and seedlings: growth promotion, methanol consumption, and localization of the methanol emission site. *J. Exp. Bot.* **57**: 4025–4032.
- 3) Tani, A., Takai, Y., Suzukawa, I., Akita, M., Murase, H., and Kimbara, K. (2012) Practical application of methanol-mediated mutualistic symbiosis between *Methylobacterium* species and a roof greening moss, *Racomitrium japonicum*. *PLoS One* **7**: e33800.
- 4) Tani, A., Ogura, Y., Hayashi, T., and Kimbara, K. (2015). Complete genome sequence of *Methylobacterium aquaticum* strain 22A, isolated from *Racomitrium japonicum* moss. *Genome Announc.* **3**: e00266-15.
- 5) Woldegiorgis, A. Z., Abate, D., Haki, G. D., and Ziegler, G. R. (2014). Antioxidant property of edible mushrooms collected from Ethiopia. *Food Chem.* **157**: 30–36.
- 6) Seebeck, F. P. (2010). In vitro reconstitution of mycobacterial ergothioneine biosynthesis. *J. Am. Chem. Soc.* **132**: 6632–6633.
- 7) Bourque, D., Pomerleau, Y., and Groleau, D. (1995). High-cell-density production of poly- $\beta$ -hydroxybutyrate (PHB) from methanol by *Methylobacterium extorquens*: production of high-molecular-mass PHB. *Appl. Microbiol. Biotechnol.* **44**: 367–376.